Molecular detection and characterization of *Cryptosporidium* spp. in the sewage-contaminated rivers entering Bandar-e Anzali Lagoon in Guilan Province, Iran

Mohammad Reza Mahmoudi^{1,2,3,⊠}

- 1. Cellular and Molecular Research Center, Faculty of Medicine, Guilan University of Medical Sciences, Rasht, Iran
- 2. Department of Parasitology, Faculty of Medicine, Guilan University of Medical Sciences, Rasht, Iran
- 3. Medical Biotechnology Research Center, Guilan University of Medical Sciences, Rasht, Iran

Date of submission: 03 Oct 2019, Date of acceptance: 22 Feb 2020

ABSTRACT

Waterborne cryptosporidiosis in river water is considered to be an important source of *Cryptosporidium* oocysts in most developing countries across the world. In the present study, 20 water samples were collected from Zarjoob and Goharrood rivers contaminated with wastewater in the province of Guilan, Iran. The samples were filtrated through a membrane filter (pore size: 1.2 µm), then the sucrose clarification and molecular genotyping methods were also carried out. In total, 12 out of 20 the samples were positive for *Cryptosporidium* species based on the 18S RNA-based polymerase chain reaction (PCR) and sequencing techniques. In addition, *C. parvum, C. muris, C. homnis*, and *C. canis* were detected in the samples studied. According to the results, the river water resources were polluted with the pathogenic species of *Cryptosporidium* with human and animal sources; this may result in an increase in the potential risk of waterborne cryptosporidiosis. Moreover, the long-term use of sewage-contaminated river water to irrigate crops and agriculture farms could be a major threat to local residents.

Keywords: Bacterial isolation, Water resources, Sewage-contaminated rivers, Guilan Province

Introduction

Cryptosporidiosis is a prevalent diarrheal disease in developing and industrialized countries. Molecular studies have helped scientists to recognize the transmission of this parasite to individuals, and the popular health significance of *Cryptosporidium* spp.¹ Currently, waterborne cryptosporidiosis is known to cause numerous diseases outbreaks mainly in the North America, United Kingdom, Japan, and Australia.²⁻⁴

Cryptosporidium oocysts have been detected in surface waters throughout Asia.⁵ A minimum of 165 water-associated outbreaks of cryptosporidiosis were describe globally since the beginning of the last century until 2004⁶ while 120 outbreaks were reported during 2004 until 2010.⁷ There are several types of *Cryptosporidium* spp., while only some species (*C. parvum* and *C. hominis*) are considered to be responsible for most of the associated human infections.⁸

In Iran, the epidemiology of *Cryptosporidium* species in environmental resources remains unclear, and constant monitoring is required in this regard. The present study aimed to provide an update on the information regarding the distribution of *Cryptosporidium* spp. in the rivers located in



[☑] Mohammad Reza Mahmoudi mrmahmoodi2002@yahoo.com

Citation: Mahmoudi M R. Molecular detection and characterization of *Cryptosporidium* spp. in the sewage-contaminated rivers entering Bandar-e Anzali Lagoon in Guilan Province, Iran. J Adv Environ Health Res 2020; 8(2): 95-99

Guilan Province, Iran and propose an approach to recognize the origin of *Cryptosporidium* oocysts in the contaminated rivers in the study area.

Materials and Methods Study area and sample collection

In total, 20 water samples were collected

from the Zarjoob and Goharrood rivers located the province of Guilan, Iran. This province is situated beside the Caspian Sea and moderate coastline has а and Mediterranean weather. The Zarjoob and Goharrood rivers are the two branches of the Sefidrood River (the second longest river in Iran), which originates in the city of Rasht and flows through the Bandar-e Anzali Lagoon. Unfortunately, these areas are threatened by numerous pollutants, such as rural, urban, and agricultural runoff.⁹ In addition, the water of these sewagecontaminated rivers is used by animals and for the irrigation of crops and agriculture farm by the local residence.

Separation of oocysts from the water samples

In this study, 20 water samples (5 L each) were collected from the depth of \sim 10-20 cm in Zarjoob and Goharrood rivers, which flow across the city of Rasht. The samples were taken in places where the rivers exited the city. Any contaminants originating from the upstream and midstream may pollute the downstream as well.

At the next step, the samples were filtrated through a cellulose acetate membrane filter (diameter: 142 mm, pore size: 1.2 μ m). The filter was washed twice with 0.1% phosphate buffered saline-Tween 80 (50 mL), and the material was concentrated via centrifugation at 3,000 rpm for 10 min. For oocyst concentration and purification, the sediment pellet was subjected to sucrose flotation¹⁰ and the supernatant was subjected to the molecular technique.

DNA extraction and PCR

DNA was extracted from the purified

oocysts using the QIAamp DNA Mini Kit as recommended previously¹¹ with some modification including ten freeze-thaw cycles (1 min in -196 °C and 1 min at 96 °C per cycle). The procedure continued until DNA extraction, which was performed based on the instructions by the manufacturer (Qiagen GmbH, Hilden, Germany). In addition, the nested PCR assay was used to amplify a 435 bp fragment of the *Cryptosporidium* spp.gene-specific 18S rRNA using the CPB-DIAGF/R and N-DIAGF2/R2 primers.^{12,13}

In accordance with previous studies, each PCR reaction was performed in 50 microliters,^{12,13} and the conditions of the first amplification were as follows: one cycle at 95 °C for 15 min, 35 cycles at 94 °C for 30 sec, 68 °C for 60 sec, and 72 °C for 30 sec, one cycle at 72 °C for 10 min, with the final hold at 4 °C. The setup of the second amplification was similar to the first procedure, with the exception of the annealing parameters, which changed to 60 °C for 60 sec. It is notable that both the positive negative controls were included in the amplifications.

The PCR products were electrophoresed on 1.5% agarose gel containing ethidium bromide (0.6 mg/mL) and imagine using a gel documentation instrument. Following that, the PCR products were sequenced using an ABI 3730XL DNA analyzer. All the sequences were edited and the genotype was identified through the comparison of the accessible *Cryptosporidium* DNA sequences in GenBank by using the Chromas software.

Results and Discussion

According to the current research, the studied rivers were contaminated with *Cryptosporidium* oocysts. In total, 12 out of 20 water samples (60%) were positive for *Cryptosporidium* spp. Some studies have detected *Cryptosporidium* oocysts in 24-100% of surface and wastewater samples collected from other regions in the world.^{14,15}

The genotyping of *Cryptosporidium* isolates contributed to the evaluation of the origin and human-infective potential of oocysts in polluted water. In the present



study, the genotyping of the isolates revealed the presence of C. parvum, C. muris, C. homnis, and C. canis. Similarly, some studies have identified the species in untreated water in Iran.¹⁶⁻¹⁸ C. hominis and C. parvum have been most common in contaminated river water as detected by PCR-based techniques worldwide. Our findings support the previous studies regarding the complexity of Cryptosporidium isolates in untreated domestic wastewater.19-21

Although C. hominis, C. parvum, and C. are all accountable for human canis infections, C. hominis and C. parvum have been shown to be more prevalent in this regard.²² In addition, C. parvum is considered to be the main species in humans with diarrhea and AIDS worldwide.²³⁻²⁶ C. hominis is a more important cause of infections than C. parvum in humans in developing nations according to genotyping researchs.²⁷⁻³² In the United Kingdom, various European regions, and New Zealand, C. parvum has been reported to be as common as C. hominis in human infections.³³⁻³⁷ The variations in spread of Cryptosporidium genotypes in human are due to the differences in the infection sources.¹

According to the literature, *C. muris* mostly infects rodents³⁸ while *C. canis* is considered to be a parasite in canines.³⁹ These species were the next most frequent genotypes detected in the present study. Furthermore, *C. canis* is responsible for human infections.^{22,39} Consistent with our findings, *C. canis* and *C. muris* have been detected in raw water and wastewater from other parts of world.^{1,35}

C. parvum has also been identified in domestic livestock and other animals, as well as humans, while C. homonis has been exclusively observed in the isolates from humans. Our findings highlighted the potential risk caused by waterborne cryptosporidiosis due to irrigation using contaminated river water. Therefore, the monitoring of Cryptosporidium in water sources is critical considering its public health implications.

Conclusion

According to the results, the river water samples were contaminated with *Cryptosporidium* oocysts. Therefore, it is hazardous for the local population to use the sewage-contaminated river water for the irrigation of crops and rice farms, and the subsequent transfer of the water to the food chain must also be prevented.

Acknowledgements

This study was supported by funding from Guilan University of Medical Sciences (grant No: 93121113).

Authors' Contributions

M. R. M: data collection, analysis, drafting and revision of the paper.

References

- 1. Feng Y, Li N, Duan L, Xiao L. *Cryptosporidium* genotype and subtype distribution in raw wastewater in Shanghai, China: Evidence for possible unique *Cryptosporidium* hominis transmission. J Clin Microbiol 2009; 47(1): 153-7.
- Imre K, Morar A, Ilie MS, Plutzer J, Imre M, Emil T, *et al.* Survey of the occurrence and human infective potential of *Giardia duodenalis* and *Cryptosporidium* spp. in wastewater and different surface water sources of western Romania. Vector Borne Zoonotic Dis 2017; 17(10): 685-91.
- 3. Efstratiou A, Ongerth J, Karanis P. Evolution of monitoring for Giardia and *Cryptosporidium* in water. Water Res 2017; 123: 96-112.
- 4. Lu P, Amburgey JE, Hill VR, Murphy JL, Schneeberger CL, Arrowood MJ, *et al.* Removals of *cryptosporidium* parvum oocysts and cryptosporidium-sized polystyrene microspheres from swimming pool water by diatomaceous earth filtration and perlite-sand filtration. J Water Health 2017; 15(3): 374-84.
- 5. Mahmoudi MR, Ongerth JE, Karanis P. *Cryptosporidium* and cryptosporidiosis: The Asian perspective. Int J Hyg Environ Health 2017; 220(7): 1098-109.
- 6. Karanis P, Kourenti C, Smith H. Waterborne transmission of protozoan parasites: A worldwide review of outbreaks and lessons learnt. J Water Health 2007; 5(1): 1–38.



- Baldursson S, Karanis P. Waterborne transmission of protozoan parasites: Review of worldwide outbreaks – an update 2004– 2010. Water Res 2011; 45(20): 6603–14.
- Plutzer J, Karanis P, Domokos K, Törökné A, Márialigeti K. Detection and characterisation of *Giardia* and *Cryptosporidium* in Hungarian raw, surface and sewagewater samples by IFT, PCR and sequence analysis of the SSUrRNA and GDH genes. Int J Hyg Environ Health 2008; 211(5-6): 524–33.
- Mahmoudi MR, Kazemi B, Haghighi A, Karanis P. Detection of *Acanthamoeba* and *Toxoplasma* in river water samples by molecular methods in Iran. Iran J Parasitol 2015; 10(2): 250-7.
- Mahmoudi MR, Ashrafi K, Abedinzadeh H, Tahvildar-Bideruni F, Haghighi A, Bandehpour M, *et al.* Development of sensitive detection of *Cryptosporidium* and *Giardia* from surface water in iran. Iran J Parasitol 2011; 6(3): 43-51.
- 11. Toledo RD, Martins FD, Ferreira FP, de Almeida JC, Ogawa L, Dos Santos HL, *et al. Cryptosporidium* spp. and *Giardia* spp. in feces and water and the associated exposure factors on dairy farms. PLoS One 2017; 12(4): e0175311.
- Matavos-Aramyan S, Moussavi M, Matavos-Aramyan H, Roozkhosh S. Cryptosporidiumcontaminated water disinfection by a novel Fenton process. Free Radic Biol Med 2017; 106: 158-67.
- 13. Gallas-Lindemann C, Sotiriadou I, Plutzer J, Noack MJ, Mahmoudi MR, Karanis P. *Giardia* and *Cryptosporidium* spp. dissemination during wastewater treatment and comparative detection via immunofluorescence assay (IFA), nested polymerase chain reaction (nested PCR) and loop mediated isothermal amplification (LAMP). Acta Trop 2016; 158: 43-51.
- Bilung LM, Tahar AS, Yunos NE, Apun K, Lim YAL, Nillian E, *et al.* Detection of *Cryptosporidium* and *Cyclospora* oocysts from environmental water for drinking and recreational activities in Sarawak, Malaysia. BioMed Research International 2017; (3): 1-9.
- 15. Xiao L, Singh A, Limor J, Graczyk TK, Gradus S, Lal A. Molecular characterization of *Cryptosporidium* oocysts in samples of raw surface water and wastewater. Appl Environ

Microbiol 2001; 67(3): 1097–101.

- 16. Mahmoudi MR, Kazemi B, Mohammadiha A, Mirzaei A, Karanis P. Detection of *Cryptosporidium* and *Giardia* (00)cysts by IFA, PCR and LAMP in surface water from Rasht, Iran. Trans R Soc Trop Med Hyg 2013; 107(8): 511-7.
- Mahmoudi MR, Nazemalhosseini-Mojarad E, Kazemi B, Haghighi A, Mirzaei A, Mohammadiha A, *et al. Cryptosporidium* genotypes and subtypes distribution in river water in Iran. J Water Health 2015; 13(2): 600-6.
- 18. Manouchehri Naeini K, Asadi M, Hashemzade Chaleshtori M. Detection and molecular characterization of *Cryptosporidium* species in recreational waters of Chaharmahal va Bakhtiyari province of Iran using nested-PCR-RFLP. Iran J Parasitol 2001; 6(1): 20–7.
- 19. Muchiri JM, Ascolillo L, Mugambi M, Mutwiri T, Ward HD, Naumova EN, *et al.* Seasonality of *Cryptosporidium* oocyst detection in surface waters of Meru, Kenya as determined by two isolation methods followed by PCR. J Water Health 2009; 7(1): 67-75.
- 20. Castro-Hermida JA, García-Presedo I, Almeida A, González-Warleta M, Correia Da Costa JM, Mezo M. Contribution of treated wastewater to the contamination of recreational river areaswith *Cryptosporidium* spp. and Giardia duodenalis. Water Res 2008; 42(13): 3528–38.
- 21. Hänninen ML, Hörman A, Rimhanen-Finne R, Vahtera H, Malmberg S, Herve S, Lahti K. Monitoring of *Cryptosporidium* and Giardia in the Vantaa river basin, southern Finland. Int J Hyg Environ Health 2005; 208(3): 163– 71.
- 22. Xiao S, Yin P, Zhang Y, Hu S. Occurrence of *Cryptosporidium* and *Giardia* and the relationship between protozoa and water quality indicators in swimming pools. Korean J Parasitol 2017; 55(2): 129-35.
- Zahedi A, Monis P, Aucote S, King B, Paparini A, Jian F, *et al.* Zoonotic *Cryptosporidium* species in animals inhabiting Sydney water catchments. PLoS One 2016; 11(12): e0168169.
- 24. Koloren Z, Ayaz E. Genotyping of *Cryptosporidium* spp. in environmental water in Turkey. Acta Parasitol 2016; 61(4): 671-9.
- 25. Pirestani M, Sadraei J, Dalimi asl A, Zavvar





M, Vaeznia H. Molecular characterization of *Cryptosporidium* isolates from human and bovine using 18 rRNA gene in Shahriar county of Tehran, Iran. Parasitol Res 2008; 103(2): 467–72.

- 26. Zavvar M, Sadraei J, Emadi H, Pirestani M. The use of a nested PCR–RFLP technique, based on the parasite's 18S ribosomal RNA, to characterize *Cryptosporidium* isolates from HIV/AIDS patients. Ann Trop Med Parasitol 2008; 102(7): 597–601.
- 27. Daniels ME, Smith WA, Schmidt WP, Clasen T, Jenkins MW. Modeling *Cryptosporidium* and *Giardia* in ground and surface water sources in rural India: Associations with latrines, livestock, damaged wells, and rainfall patterns. Environ Sci Technol 2016; 50(14): 7498-507.
- 28. Maciel PMF, Sabogal-Paz LP. Removal of *Giardia* spp. and *Cryptosporidium* spp. from water supply with high turbidity: Analytical challenges and perspectives. J Water Health 2016; 14(3): 369-78.
- 29. Koehler AV, Haydon SR, Jex AR, Gasser RB. *Cryptosporidium* and *Giardia* taxa in faecal samples from animals in catchments supplying the city of Melbourne with drinking water (2011 to 2015). Parasit Vectors 2016; 9(1): 315.
- 30. Chuah CJ, Mukhaidin N, Choy SH, Smith GJD, Mendenhall IH, Lim YAL, *et al.* Prevalence of *Cryptosporidium* and *Giardia* in the water resources of the Kuang River catchment, Northern Thailand. Sci Total Environ 2016; 562: 701-13.
- 31. 31. Sterk A, Schijven J, de Roda Husman AM, de Nijs T. Effect of climate change on runoff of *Campylobacter* and *Cryptosporidium* from land to surface water. Water Res 2016; 95: 90-102.
- 32. Kumar T, Abd Majid MA, Onichandran S, Jaturas N, Andiappan H, Salibay CC, *et al.* Presence of *Cryptosporidium* parvum and *Giardia* lamblia in water samples from

Southeast Asia: Towards an integrated water detection system. Infect Dis Poverty 2016; 5: 3.

- 33. Headd B, Bradford SA. Use of aerobic spores as a surrogate for *Cryptosporidium* oocysts in drinking water supplies. Water Res 2016; 90: 185-202.
- 34. Hawash Y, Ghonaim M, Hussein Y, Alhazmi A, Alturkistani A. Identification of Giardia lamblia and the human infectious-species of *Cryptosporidium* in drinking water resources in Western Saudi Arabia by nested-PCR assays. Trop Biomed 2015; 32(2): 216-24.
- 35. Zahedi A, Paparini A, Jian F, Robertson I, Ryan U. Public health significance of zoonotic *Cryptosporidium* species in wildlife: Critical insights into better drinking water management. Int J Parasitol Parasites Wildl 2016; 5(1): 88-109.
- 36. Mesdaghinia A, Younesian M, Nasseri S, Nabizadeh Nodehi R, Hadi M. A bibliometric and trend analysis on the water-related risk assessment studies for *Cryptosporidium* pathogen. Iran J Parasitol 2015; 10(3): 338-50.
- 37. Murphy HM, Thomas MK, Schmidt PJ, Medeiros DT, McFadyen S, Pintar KDM. Estimating the burden of acute gastrointestinal illness due to *Giardia*, *Cryptosporidium*, *Campylobacter*, *E. coli O157* and norovirus associated with private wells and small water systems in Canada. Epidemiol Infect 2016; 144(7): 1355-70.
- Abeledo-Lameiro MJ, Ares-Mazas E, Gomez-Couso H. Evaluation of solar photocatalysis using TiO₂ slurry in the inactivation of *Cryptosporidium parvum* oocysts in water. J Photochem Photobiol B 2016; 163: 92-9.
- 39. Xiao L, Cama VA, Cabrera L, Ortega Y, Pearson J, Gilman RH. Possible transmission of *Cryptosporidium canis* among children and a dog in a household. J Clin Microbiol 2007; 45(6): 2014–6.

